

PIPERIDIDES AND OTHER AMIDES FROM *ACHILLEA* SPECIES*

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Key Word Index—*Achillea spinulifolia*; *A. grandifolia*; *A. lycaonica*; *A. tomentosa*; Compositae; Anthemideae; amides; piperidides; pyrrolidides; acetylenes.

Abstract—The investigation of the roots of four *Achillea* species afforded, in addition to known ones eight new amides most of them being piperidides. The structures were elucidated by spectroscopic methods. The chemotaxonomic importance of the amides is discussed briefly.

INTRODUCTION

So far the chemical investigations of 13 *Achillea* species have shown that acetylenic compounds with an amide grouping are characteristic for this genus [1-4]. These compounds have also been isolated from members of the genera *Anacyclus* [5], *Otanthus* [5] and *Leucocyclus* [4] which are probably closely related to *Achillea*. We have now investigated three further species and re-investigated *A. tomentosa*. Several known and eight new amides were isolated.

RESULTS AND DISCUSSION


The roots of *Achillea spinulifolia* Fenzl. ex Boiss. afforded the *iso*-butyl-amides 8 [1], 9 [6] and 12 [7] as well as two further ones, the acetylenic piperidides 1 and 3. The structure of 1 followed from the spectroscopic data. The UV maximum indicated the presence of a dienamide, which was established by the ¹H NMR data (Table 1) and which further indicated that this amide was a piperidide as followed from the typical multiplets at δ 3.79, 3.67, 1.64 and 1.56. The nature of the unconjugated side chain was also deduced from the ¹H NMR data. The presence of a *cis*-double bond followed from the coupling of the olefinic signals, while the propyl end group was deduced by spin decoupling. Also the fragmentation pattern in the mass spectrum supported the structure (see Experimental). This was also true for the structure of 3, where the fragments at m/z 101 [$\text{Me}(\text{C}\equiv\text{C})_3\text{CH}_2$]⁺ and $[M - 101]^+$ clearly indicated the nature of the unsaturation. The ¹H NMR data (Table 1) in part were similar to those of 1. However, the H-6 and H-7 signals were an overlapping multiplet

and the H-14 signal was a sharp singlet, indicating that more than two triple bonds must be present. The roots of *A. grandifolia* Friv. afforded two amides, the enediyne 10 and a second diyne, which could be separated from 10 only with difficulty. Its molecular formula showed that this amide had two hydrogens less than 10. The UV spectrum was similar to that of an enediyne triene carboxylate [8], while the ¹H NMR data (Table 1) led to structure 11, as was deduced from spin decoupling experiments. Starting with the H-2 signal the H-3 signal could be assigned, which was partly obscured by the chloroform signal. However, in C₆D₆ this signal could be observed. Further decoupling allowed the assignment of H-4 through H-7, while the nature of the end group directly followed from the typical signals of a propenyl group linked to an acetylenic bond. The configuration of the double bond followed from the coupling constants observed. Compound 11 is the first fully conjugated C₁₄-amide.

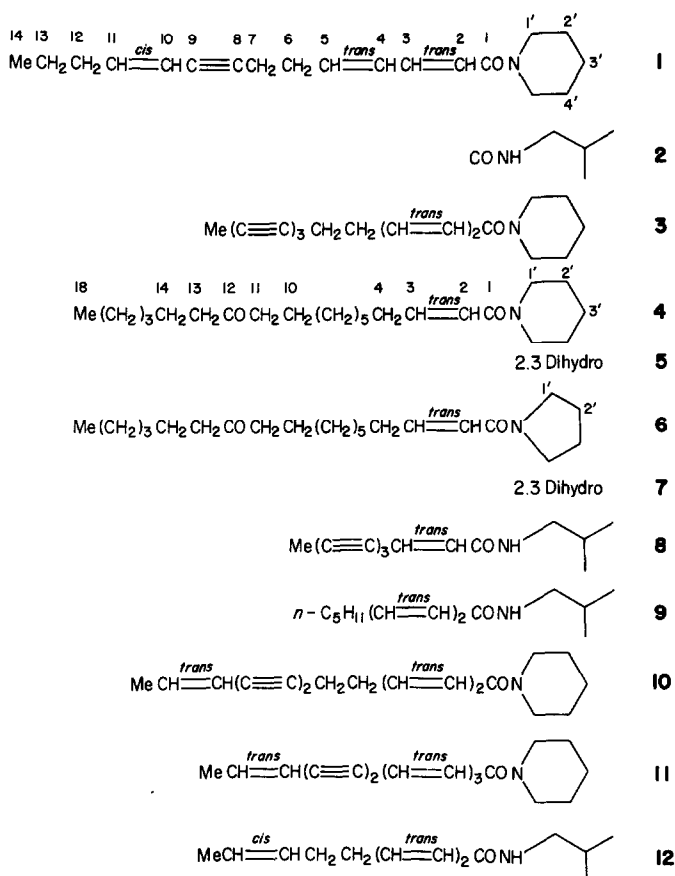
The roots of *A. lycaonica* Boiss. et Heldr. afforded two amide fractions, which both crystallized easily. The mass spectra as well as the ¹H NMR spectra, however, showed that both were mixtures of two amides, differing by two hydrogens. Inspection of the ¹H NMR data led to the proposal each time that we were dealing with an α,β -unsaturated and the corresponding saturated amide. Consequently a separation with silver nitrate impregnated Si gel was possible to produce four amides with the molecular formulae C₂₃H₄₃O₂, C₂₃H₄₁NO₂, C₂₂H₄₁O₂ and C₂₁H₃₉NO₂. The ¹H NMR data (Table 1) indicated that the first pair were piperidides with and without a 2,3-*trans*-double bond, while the second one were pyrrolidides also differing in saturation. The nature of the second oxygen function followed from the IR spectra, while its position was deduced from the mass spectra, which in all cases showed elimination of Me⁺, Et⁺, C₃H₇⁺, C₄H₉⁺. The last fragment showed loss of a

*Part 261 in the series "Polyacetylenic Compounds". For Part 260 see Greger, H., Grenz, M. and Bohlmann, F. (1981) *Phytochemistry* 20, 2579.

Table 1. ¹H NMR spectral data of compounds 1-7 and 11 (400 MHz, CDCl₃, TMS as int. standard)

	1	2	3	4	5	6	7	11	
H-2	6.28 <i>br d</i>	5.78 <i>br d</i>	6.31 <i>br d</i>	6.24 <i>dt</i>	2.30 <i>t</i>	6.08 <i>dt</i>	2.25 <i>t</i>	6.43 <i>d</i>	
H-3	7.22 <i>dd</i>	7.19 <i>dd</i>	7.21 <i>dd</i>	6.82 <i>dt</i>	1.54 <i>m</i>	6.91 <i>de</i>	1.63 <i>m</i>	7.27 <i>dd</i>	
H-4	6.24 <i>br dd</i>	6.20 <i>br dd</i>	6.23 <i>br dd</i>	2.18 <i>ddt</i>	1.29 <i>m</i>	2.19 <i>ddt</i>	1.27 <i>m</i>	6.73 <i>dd</i>	
H-5	6.08 <i>br dt</i>	6.10 <i>br dt</i>	6.01 <i>br dt</i>	1.20 <i>m</i>		1.27 <i>m</i>		6.38 <i>dd</i>	6.52 <i>dd</i>
H-6	2.40 <i>br dt</i>	2.40 <i>br dt</i>	2.40 <i>m</i>					5.80 <i>br d</i>	
H-7	2.46 <i>br t</i>	2.47 <i>br dt</i>	—	—	—	—	—	—	
H-8, H-9	—	—	—	—	—	—	—	—	
H-10	5.43 <i>dt</i>	5.43 <i>br d</i>	—	1.57 <i>m</i>	1.54 <i>m</i>	1.55 <i>m</i>	1.55 <i>m</i>	—	
H-11	5.83 <i>dt</i>	5.83 <i>dt</i>	—	2.38 <i>t</i>	2.38 <i>t</i>	2.38 <i>t</i>	2.38 <i>t</i>	—	
H-12	2.25 <i>ddt</i>	2.24 <i>ddt</i>	—	—	—	—	—	—	
H-13	1.42 <i>tq</i>	1.41 <i>tq</i>	—	2.38 <i>t</i>	2.38 <i>t</i>	2.38 <i>t</i>	2.38 <i>t</i>	—	
H-14	0.91 <i>t</i>	0.91 <i>t</i>	1.95 <i>s</i>	1.57 <i>m</i>	1.57 <i>m</i>	1.55 <i>m</i>	1.53 <i>m</i>	—	
H-15-H-17	—	—	—	1.29 <i>m</i>	1.29 <i>m</i>	1.29 <i>m</i>	1.29 <i>m</i>	5.60 <i>br d</i> , 6.33 <i>tq</i>	
H-18	—	—	—	0.88 <i>t</i>	0.88 <i>t</i>	0.87 <i>t</i>	0.87 <i>t</i>	3.61 <i>m</i>	
	3.79 <i>m</i>	—	3.61 <i>m</i>	3.59 <i>m</i>	3.53 <i>t</i>	3.52 <i>t</i>	3.46 <i>t</i>	3.61 <i>m</i>	
	3.67 <i>m</i>	—	3.49 <i>m</i>	3.48 <i>m</i>	3.38 <i>t</i>	—	3.41 <i>t</i>	3.49 <i>m</i>	
	1.56 <i>m</i>	—	1.57 <i>m</i>	1.57 <i>m</i>	1.54 <i>m</i>	1.96 <i>tt</i>	1.84 <i>tt</i>	1.57 <i>m</i>	
	1.64 <i>m</i>	—	1.64 <i>m</i>	1.65 <i>m</i>	1.61 <i>m</i>	1.82 <i>tt</i>	1.94 <i>tt</i>	1.65 <i>m</i>	

J(Hz): Compounds 1-3: 2,3 = 4,5 = 15; 3,4 = 10.5; 5,6 = 6,7 = 7; compounds 1 and 2: 10,11 = 10; 10,12 = 1.5; 11,12 = 13,14 = 7; compounds 4-7: 10,11 = 13,14 = 17,18 = 7; compounds 4 and 6: 2,3 = 15; 2,4 = 1.5; 3,4 = 4,5 = 7; compounds 5 and 7: 2,3 = 7; compound 11: 2,3 = 4.5 = 6,7 = 16,17 = 15; 3,4 = 5,6 = 10; 16,18 = 1.5; 17,18 = 7; NHtBu: 1', NH = 1',2' = 2',3' = 2',4' = 7; pyrrolidides 1',2' = 7.



Scheme 1.

carbonyl, indicating that the keto group could only be at C-12. Furthermore the corresponding peaks from a McLafferty fragmentation were visible (m/z 295, 293, 281 and 279 respectively). The other fragments were formed by splitting all single bonds of C-1 through C-11, though some fragments were more intensive indicating some preferences (Scheme 1). As all compounds also showed a prominent fragment, due to the corresponding acetyl amide, migration of the double bond must also have taken place which probably led to the favoured formation of some ions. All data were in agreement with structures 4–7. The clear difference in polarity of the pairs 4, 5 and 6, 7 is interesting. The ¹H NMR data of H-2 and H-3, as well as those of the amide part, were also typically different. We have named the acid corresponding to 5 and 7 lycaonic acid.

A re-investigation of the extract of the roots of *A. tomentosa* L. afforded a further compound, the isobutylamide 2. Its structure clearly followed from the ¹H NMR data (Table 1), which were similar to those of 1, except for the amide part. These results again showed that different types of amides are typical for *Achillea* species. Further investigations are required to show to what extent these compounds may contribute to a species grouping within the genus.

EXPERIMENTAL

The roots were extracted with Et₂O–petrol (1:2) and the

resulting extracts were separated first by CC (Si gel) and the polar fractions (Et₂O–petrol, 1:1 and Et₂O) further by repeated TLC (Si gel and partly AgNO₃–Si gel). Specimen vouchers are deposited in the Herbarium of the Institute of Botany, University of Vienna.

Achillea lycaonica (voucher A-1542). The roots (238 g) afforded 40 mg 3 and 4 (ca 1:1) as well as 40 mg 5 and 6 (ca 1:1). Separation on AgNO₃–Si gel (Et₂O–petrol, 7:3, ×3) gave the pure compounds.

Achillea spinulifolia (voucher A-1540). The roots (83 g) afforded 75 mg 1, 8.4 mg 3, 20 mg 8, 95 mg 9 and 10 mg 12.

Achillea grandifolia (voucher A-1443). The roots (130 g) afforded 5 mg 10 and 11 (ca 3:2), which could be separated by TLC on AgNO₃–Si gel, but only 1 mg pure 11 was obtained.

Achillea tomentosa (voucher A-1602). The roots (31 g) afforded in addition to the amides reported earlier [4] 5 mg 2 (AgNO₃–Si gel, Et₂O–petrol, 4:1).

Tetradeca-2t, 4t, 10c-trien-8-ynoic-piperidide (1). Colourless gum, IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm⁻¹: 2230 (C≡C), 1655, 1630, 1605 (C=CCON<); UV $\lambda_{\text{max}}^{\text{Et}_2\text{O}}$ nm: 257; MS m/z (rel. int.): 285.209 [M]⁺ (11)(C₁₉H₂₇NO), 284 [M–H]⁺ (12), 270 [M–Me]⁺ (3), 256 [M–Et]⁺ (6), 242 [M–C₃H₇]⁺ (2), 179 [M–C₄H₉]⁺ (41) (McLafferty), 178 [M–C₃H₇CH=CHCCH₂]⁺ (30), 164 [179–Me]⁺ (14), 112 [CONC₃H₁₀]⁺ (100), 84 [112–CO]⁺ (93).

Tetradeca-2t, 4t, 10c-trien-8-ynoic-iso-butylamide (2). Colourless oil, IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm⁻¹: 3440, 1670, 1620 (CH=CHCONH); UV $\lambda_{\text{max}}^{\text{Et}_2\text{O}}$ nm: 252; MS m/z (rel. int.):

273.204 [M]⁺ (6) (C₁₈H₂₇NO), 272 [M - H]⁺ (5), 201 [M - NHR]⁺ (5), 173 [201 - CO]⁺ (20), 57 [C₄H₉]⁺ (100).

Tetradeca - 2t, 4t - diene - 8,18,12 - triynoic piperidide (3). Colourless crystals, mp 115°; IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm⁻¹: 2230 (C≡C), 1650, 1625, 1605 (C=CCOH); UV $\lambda_{\text{max}}^{\text{Et}_2\text{O}}$ nm: 253; MS *m/z* (rel. int.): 279.162 [M]⁺ (84) (C₁₉H₂₁NO), 264 [M - Me]⁺ (2), 205 [M - NC₃H₁₀]⁺ (17), 178 [M - Me (C≡C)₃CH₂]⁺ (41), 101 [Me (C≡C)₃CH₂]⁺ (81), 84 [NC₃H₁₀]⁺ (100).

2,3-Dehydrolycaonic acid piperidide (4). Colourless crystals, mp 43°; IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm⁻¹: 1715 (C=O), 1660, 1620 (C=CCON); MS *m/z* (rel. int.): 363.314 [M]⁺ (2) (C₂₃H₄₁NO₂), 348 [M - Me]⁺ (0.3), 334 [M - Et]⁺ (0.5), 320 [M - C₃H₇]⁺ (0.5), 306 [M - C₄H₉]⁺ (3), 293 [M - C₅H₁₀]⁺ (2) (McLafferty), 278 [M - C₆H₁₃]⁺ (2), 250 [278 - CO]⁺ (6), 236 [M - CH₂COC₆H₁₃]⁺ (14), 222 (3), 208 (4), 194 (4), 180 (7), 166 (20), 138 [CH=CHCONC₅H₁₀]⁺ (100), 127 [MeCON]⁺ (12), 112 [138 - C₂H₂]⁺ (10), 84 [NC₃H₁₀]⁺ (24).

Lycaonic acid piperidide (5). Colourless crystals, mp 41°; IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm⁻¹: 1715 (C=O), 1645 (CON); MS *m/z* (rel. int.):

365.329 [M]⁺ (4) (C₂₃H₄₃NO₂), 350 [M - Me]⁺ (0.2), 336 [M - Et]⁺ (0.2), 322 [M - C₃H₇]⁺ (0.2), 308 [M - C₄H₉]⁺ (2), 295 [M - C₅H₁₀]⁺ (1.5) (McLafferty), 280 [M - C₆H₁₃]⁺ (4), 252 [280 - CO]⁺ (6), 238 [M - CH₂COC₆H₁₃]⁺ (27), 224 (2), 210 (2), 196 (4), 182 (6), 168 (4), 154 (8), 140 (27), 127 [MeCONC₅H₁₀]⁺ (100), 112 [CONC₅H₁₀]⁺ (16), 84 [NC₃H₁₀]⁺ (21).

2,3-Dehydrolycaonic acid pyrrolidide (6). Colourless crystals, mp 64°; IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm⁻¹: 1715 (C=O), 1660, 1615 (C=CCON); MS *m/z* (rel. int.): 349.297 [M]⁺ (18) (C₂₂H₃₉NO₂), 306 [M - C₃H₇]⁺ (1), 292 [M - C₄H₉]⁺ (8), 279 [M - C₅H₁₀]⁺ (4) (McLafferty), 264 [M - C₄H₉]⁺ (6), 236

[252 - CO]⁺ (34), 222 [M - CH₂COC₆H₁₃]⁺ (95), 208 (7), 194 (8), 180 (8), 166 (10), 102 (100), 124 (60), 113 [MeCONC₅H₁₀]⁺ (83), 98 [124 - C₂H₂]⁺ (30), 70 [98 - CO]⁺ (72).

Lycaonic acid pyrrolidide (7). Colourless crystals, mp 44°; IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm⁻¹: 1715 (C=O), 1645 (CON); MS *m/z* (rel. int.):

351.314 [M]⁺ (2) (C₂₂H₄₁NO₂), 336 [M - Me]⁺ (0.3), 294 [M - C₄H₉]⁺ (2), 281 [M - C₅H₁₀]⁺ (2) (McLafferty), 266 [M - C₆H₁₃]⁺ (3), 238 [266 - CO]⁺ (4), 224 (18), 210 (2), 196 (4), 182 (4), 168 (4), 126 (20), 113 (100), 70 (30).

Tetradeca - 2t,4t,6t,12t - tetraene - 8,10 - diynoic piperidide (11). Unstable, yellow gum, UV $\nu_{\text{max}}^{\text{Et}_2\text{O}}$ nm: 372, 348, 266; IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm⁻¹: 2220 (C≡C), 1650 (CON); MS *m/z* (rel. int.): 279.162 [M]⁺ (19) (C₁₉H₂₁MO), 278 [M - H]⁺ (9), 264 [M - Me]⁺ (6), 195 [M - NC₃H₁₀]⁺ (11), 167 [195 - CO]⁺ (24), 165 [195 - H₂]⁺ (60), 84 [NC₃H₁₀]⁺ (100).

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